

**Materials List for:****Visualizing RNA Localization in *Xenopus* Oocytes**James A. Gagnon<sup>1</sup>, Kimberly L. Mowry<sup>1</sup><sup>1</sup>Department of Molecular Biology, Cell Biology, and Biochemistry, Brown UniversityCorrespondence to: Kimberly L. Mowry at [Kimberly\\_Mowry@brown.edu](mailto:Kimberly_Mowry@brown.edu)URL: <https://www.jove.com/video/1704>

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**Materials**

<b>10X Tx buffer</b>
<ul style="list-style-type: none"><li>• 60 mM MgCl<sub>2</sub></li><li>• 400 mM Tris-HCl (pH 7.5)</li><li>• 20 mM spermidine-HCl</li></ul>
<b>20x cap/NTP mix</b>
<ul style="list-style-type: none"><li>• 10 mM CTP</li><li>• 10 mM ATP</li><li>• 9 mM UTP</li><li>• 2 mM GTP</li><li>• 20 mM G(PPP)G Cap Analog (New England Biolabs)</li></ul>
<b>G-50 column</b>
<ul style="list-style-type: none"><li>• Hydrate 5 g Sephadex G-50 beads (Sigma Aldrich) in 100 ml deionized H<sub>2</sub>O. DEPC-treat for 30 min. and autoclave. Store incomplete stock at room temperature. Before use, add the following RNase-free solutions:<ul style="list-style-type: none"><li>• 0.5 ml 0.2 M EDTA</li><li>• 1 ml 1 M Tris pH 8.0</li><li>• 0.5 ml 20% SDS</li></ul></li><li>• Store complete G-50 solution at 4 ° C.</li><li>• Remove and discard the plunger from a 3 ml syringe (BD Biosciences) and place the barrel of the syringe into a 15 ml conical tube (Corning). Plug the syringe with a small amount of glass wool (a plug about half the size of a penny).</li><li>• Swirl complete G-50 solution to resuspend beads.</li><li>• Add 2 ml G-50 solution to the empty column.</li><li>• Spin for 1 minute at 1,000 x g in benchtop centrifuge.</li><li>• Add 200 µl DEPC-treated deionized H<sub>2</sub>O to each column. Spin.</li><li>• Repeat wash twice more for a total of three washes.</li><li>• Remove syringe barrel to a fresh 15 ml conical tube.</li></ul>
<b>Collagenase solution</b>
<ul style="list-style-type: none"><li>• 75 mg collagenase from <i>Clostridium histolyticum</i> (Sigma Aldrich)</li><li>• 25 ml 0.1 M KPO<sub>3</sub><sup>+</sup> (pH 7.4)</li></ul>
<b>MBSH buffer</b>
<ul style="list-style-type: none"><li>• 88 mM NaCl</li><li>• 1 mM KCl</li><li>• 2.4 mM NaHCO<sub>3</sub></li><li>• 0.82 mM MgSO<sub>4</sub> X 7H<sub>2</sub>O</li><li>• 0.33 mM Ca(NO<sub>3</sub>)<sub>2</sub> X 4H<sub>2</sub>O</li><li>• 0.41 mM CaCl<sub>2</sub> X 6H<sub>2</sub>O</li><li>• 10 mM HEPES (pH 7.6)</li></ul>
<b>Oocyte Culture Medium</b>
<ul style="list-style-type: none"><li>• 50% L15 medium</li><li>• 15 mM HEPES (pH 7.6)</li><li>• 1 mg/ml insulin</li></ul>

- 100 mg/ml gentamicin
- 50 U/ml nystatin
- 50 U/ml penicillin
- 50 mg/ml streptomycin

**MEMFA solution**

- 0.1 M MOPS (pH 7.4)
- 2 mM EGTA
- 1 mM MgSO<sub>4</sub>
- 3.7% formaldehyde

**Computing RNA yield**

- Determine CPM in "input" and "incorporated" samples using a standard scintillation counter.
- incorporation = ("incorporated") / (10 x "input")
- Typical incorporation values range between ~0.03 and 0.10.
- Maximum theoretical yields for different polymerases: T7, T3, SP6 - 2.64 µg
- Reaction yield in µg = (maximum yield of polymerase used) X (incorporation)
- Dilute RNA to 50 nM = (µg RNA) / 320 / (length of RNA in bases) / (5X10<sup>-8</sup>)
- The reaction usually yields ~50-100 µl of RNA at 50 nM.