**Recipes:**

**20% dextrose solution:**

**For one liter of 20% dextrose:** Dissolve 200g of dextrose in 600mL of MilliQ water. Once all the dextrose has dissolved, bring the solution up to 1000mL with MilliQ water and filter sterilize using a 0.45μm or 0.22μm membrane.

**Complete dropout powder mix:** In a 500mL beaker, add together 1.25g adenine, 0.9g arginine, 3.0g aspartate, 3.0g glutamate, 0.9g lysine, 0.6g methionine, 1.5g phenylalanine, 11.25g serine, 6.0g threonine, 0.9g tyrosine, 4.5g valine, 1.2g alanine, 1.2g asparagine, 1.2g cysteine, 1.2g glutamine, 1.2g glycine, 1.2g isoleucine, 1.2g proline, 0.6g histidine, 1.8g leucine, 1.2g tryptophan, and 0.6g uracil. Once all powders have been deposited in the beaker, mix thoroughly with a spatula. In small increments, transfer portions of the mixed powder to a mortar, and gently crush with a pestle. Transfer the crushed powder to a new 500mL beaker. Once all powder has been crushed, mix again thoroughly and store in 50mL conical tubes at room temperature.

**Synthetic-Complete (SC) media:** For one liter of SC media, dissolve the following in 600mL of MilliQ water: 10g succinic acid, 6g sodium hydroxide, 5g ammonium sulfate, 1.7g yeast nitrogen base without ammonium sulfate and amino acids, and 1.3g complete dropout powder mix (see recipe for details). Once all components are dissolved, bring the solution up to 900mL wilh MilliQ water and autoclave to sterilize. After the media has cooled, add 100mL of filter-sterilized 20% dextrose to the media (final dextrose concentration = 2%).

**p-anisaldehyde reagent:** For 100mL of p-anisaldehyde spray reagent: Add 500μL of p-anisaldehyde to 10mL of glacial acetic acid. Once mixed, add 85mL of 200 proof absolute ethanol and 5mL of concentrated sulfuric acid.