# Coverslip cleaning for live cell FCS experiments

## Goal

Cleaning of glass coverslips to minimize background signal

## Comments

* Cleaning procedure must be performed under the hood
* Use safety gloves when handling chloroform
* Reuse chloroform and NaOH solution
* Prepare 5 M NaOH solution on ice
* Materials: ultrasonic bath, 2 glass containers, metal rack, small beaker, funnel, 25 ml glass pipet, glass petri dish
* Discard used chemicals according to the local regulations.

## Cleaning Protocol

* Arrange single coverslips in metal rack

|  |  |
| --- | --- |
| C:\Users\User\AppData\Local\Microsoft\Windows\INetCache\Content.Word\P1112673.jpg | C:\Users\User\AppData\Local\Microsoft\Windows\INetCache\Content.Word\P1112677.jpg |

* Place glass container in ultrasonic bath, fill bath with deionized water
* Hang the rack into the glass container and fix with the loops

|  |
| --- |
| C:\Users\User\AppData\Local\Microsoft\Windows\INetCache\Content.Word\P1112681.jpg |

* **Fill the glass container with chloroform and sonicate for 1 hour**
* **Place the rack in an empty glass container and let it dry**
* Put on **safety gloves**!
* Pour the chloroform back into the bottle via a funnel by first scooping it with a beaker, then pipette the rest with a 25 ml glass pipette. Do **NOT** pour it directly out of the glass container! It will spill badly!
* Reinstall the rack in the glass container in the ultrasonic bath.
* **Fill the glass container with 5 M NaOH solution and sonicate for 1 hour**
* **Wash three times in ddH2O in a second glass container**
* **Dry glass coverslips**
* **Store in 100 % ethanol in a glass petri dish**

|  |
| --- |
| **C:\Users\User\AppData\Local\Microsoft\Windows\INetCache\Content.Word\P1112691.jpg** |

* Pour the NaOH back into the bottle as described before. Clean pipette by pipetting some ddH2O.