

TSE Immunohistochemical Test Record

Procedure	Time	Comments	Signature
1 Deparaffinization 1.1. Xylene <input type="checkbox"/> 1.2. Xylene <input type="checkbox"/> 1.3. Absolute Ethanol <input type="checkbox"/> 1.4. Absolute Ethanol <input type="checkbox"/> Air dry the slides 1.5. Ethanol 90% <input type="checkbox"/> 1.6. Ethanol 70% <input type="checkbox"/> 1.7 Ethanol 50% <input type="checkbox"/>	3 min each 3 min each 1 min each		
2 Formic acid 98 %	30 min		
3 Rinse under running water	5 min		
4 Rinse with distilled water <input type="checkbox"/> <input type="checkbox"/>			
5 Autoclaving 121°C Citrate buffer pH..... date.....	30min	Dilute stock solution Adjust Ph	
6 Cool the slides in water	5 min		
7 Hydrogen Peroxide 3% in methanol	See TSEIHC worksheet	Prepare before use	
8 Rinse under running water			
9 Immersion in TBS TBS date.....		Dilute stock solution	
10 Mount slides on Coverplate in TBS or in wet chamber			
11 Incubation with 20% normal horse serum Note: Do not rinse	See TSEIHC worksheet	Prepare before use	
12 Primary antibody		Prepare before use	
13 Wash with TBS			
14 Biotinylated Secondary Antibody		Prepare before use	
15 Wash with TBS			
16 ABC/HRP Complex		Prepare 30 min before use	
17 Wash with TBS			
18 Chromogen: DAB <input type="checkbox"/> AEC <input type="checkbox"/>		Prepare immediately before use	
19 Rinse with distilled water and remove from coverplate			
20 Counterstaining Erlich's Hematoxylin	1 min		
21 Rinse in running water until the tissues turn blue	10 min		
22 Dehydration 22.1. Ethanol 90% <input type="checkbox"/> 22.2. Absolute Ethanol <input type="checkbox"/> 22.3 Xylene <input type="checkbox"/>	1 min each		
23 Mounting Entellan <input type="checkbox"/> Glycergel <input type="checkbox"/>			
Apply 200 µL of each reagent per slide and 400 µL for the chromogen			