**Abstract**

Exercise is a stress to the pulmonary vasculature. With incremental exercise, the pulmonary diffusion capacity (DL\textsubscript{CO}) must increase to meet the increased oxygen demand; otherwise, a diffusion limitation may occur. The increase in DL\textsubscript{CO} with exercise is due to increased capillary blood volume (Vc) and membrane diffusing capacity (Dm). Vc and Dm increase secondary to the recruitment and distension of pulmonary capillaries, increasing the surface area for gas exchange and decreasing pulmonary vascular resistance, thereby attenuating the increase in pulmonary arterial pressure. At the same time, the recruitment of intrapulmonary arteriovenous anastomoses (IPAVA) during exercise may contribute to gas exchange impairment and/or prevent large increases in pulmonary artery pressure.

We describe two techniques to evaluate pulmonary diffusion and circulation at rest and during exercise. The first technique uses multiple-breath holds to determine Vc and Dm at rest and during exercise. Additionally, echocardiography with intravenous agitated saline contrast is used to assess IPAVAs recruitment.

Representative data showed that the DL\textsubscript{CO}, Vc, and Dm increased with exercise intensity. Echocardiographic data showed no IPAVA recruitment at rest, while contrast bubbles were seen in the left ventricle with exercise, suggesting exercise-induced IPAVA recruitment.

The evaluation of pulmonary capillary blood volume, membrane diffusing capacity, and IPAVA recruitment using echocardiographic methods is useful to characterize the ability of the lung vasculature to adapt to the stress of exercise in health as well as in diseased groups, such as those with pulmonary arterial hypertension and chronic obstructive pulmonary disease.

**Video Link**

The video component of this article can be found at https://www.jove.com/video/54949/

**Introduction**

During exercise, cardiac output can increase up to six-fold above resting values\textsuperscript{1}. Given that the lungs are the only organ to receive 100% of the cardiac output, exercise presents a considerable stress to the pulmonary system. With incremental exercise, pulmonary diffusion capacity (DL\textsubscript{CO}) must increase to meet the increased oxygen demand\textsuperscript{2}. From rest to peak exercise, DL\textsubscript{CO} can increase to up to 150% of resting values without reaching an upper limit with respect to cardiac output\textsuperscript{3,4,5}. The increase in diffusion capacity occurs as a result of increases in membrane diffusing capacity (Dm) and capillary blood volume (Vc), secondary to the recruitment and distension of pulmonary capillaries\textsuperscript{6}.

Roughton and Forster (1957) developed a technique to partition Dm and Vc\textsuperscript{7} by modulating the fraction of inspired oxygen (F\textsubscript{O\textsubscript{2}}) DL\textsubscript{CO} breath holds to determine Vc and Dm at rest and during exercise. Additionally, echocardiography with intravenous agitated saline contrast is used to assess IPAVAs recruitment.

Roughton and Forster (1957) developed a technique to partition Dm and Vc\textsuperscript{7} by modulating the fraction of inspired oxygen (F\textsubscript{O\textsubscript{2}}) DL\textsubscript{CO} during a standard diffusion capacity for carbon monoxide test (DL\textsubscript{CO}). Oxygen and carbon monoxide (CO) competitively bind to heme sites on hemoglobin, such that increasing F\textsubscript{O\textsubscript{2}} will decrease the DL\textsubscript{CO}\textsuperscript{8,9}. By modulating the F\textsubscript{O\textsubscript{2}} during a standard DL\textsubscript{CO} maneuver, this relationship can be exploited to measure Vc and Dm\textsuperscript{7}. We have recently adapted this technique to be used during exercise\textsuperscript{5}. Similar to previous work, we have found that DL\textsubscript{CO} continuously increases up to peak exercise secondarily to increases in both Vc and Dm\textsuperscript{7}. Interestingly, we have found that in endurance-trained athletes who have a greater oxygen consumption and thus a greater need for diffusing capacity, there is an increase in the DL\textsubscript{CO} at peak exercise, secondary to an increased Dm, and not Vc, suggesting a potential adaptation in the pulmonary membrane of the athlete\textsuperscript{5}.

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The increases in VC and Dm during exercise are accomplished by an increase in pulmonary artery pressure, which results in the recruitment and distension of pulmonary capillaries previously hypo-perfused at rest. This results in an increase in the cross-sectional area of the pulmonary capillary network, thereby decreasing pulmonary vascular resistance and attenuating the increase in pulmonary artery pressure.

Studies using agitated saline contrast echocardiography have shown evidence of intrapulmonary arteriovenous anastomoses (IPAVA) recruitment during exercise. The significance of IPAVA recruitment is not yet clear, and while some studies suggest that they may contribute to gas exchange impairment and may serve to unload the right ventricle, the topic remains controversial. Further, while the exact mechanism of IPAVA recruitment is not known, it has been found that increasing cardiac output, as well as exogenous dopamine, causes IPAVA recruitment at rest. An acutely increasing pulmonary artery pressure or dopamine blockade does not appear to significantly affect IPAVA recruitment during exercise. There is speculation that these larger-diameter IPAVA vessels may help to protect the pulmonary capillaries from the large increases in pulmonary artery pressure by reducing pulmonary vascular resistance.

When combined with the evaluation of VC and Dm, agitated saline contrast echocardiography is a valuable tool to examine the adaptation of the pulmonary circulation to the stress of exercise.

### Protocol

This protocol follows the guidelines of the human research ethics board at the University of Alberta and conforms to the standards set by the latest revision of the Declaration of Helsinki.

#### 1. Graded Exercise Test (VO₂peak)

1. Obtain written, informed consent from the subject. Have the subject read and answer the questions listed on the Physical Activity Readiness Questionnaire+ (PAR-Q+) to determine their readiness for exercise.
2. Adjust the seat height of the cycle ergometer in accordance to subject preference. Place four electrocardiogram (ECG) electrodes on the back of the patient according to standard 3-lead ECG placement, with modified limb leads to measure the heart rate (HR)
3. Insert the mouthpiece into the subject's mouth to measure the exhaled gas and ventilation throughout the test using a metabolic measurement system
4. Following 2 min of collection of baseline data, instruct the subject to start cycling with an initial workload of 50 watt, to maintain a consistent rate (HR), and end tidal CO₂ (PETCO₂).
5. For resting measurements, have the subject seated upright, with both feet flat on the floor. For exercise trials, ensure that the subject is in a back of the patient according to standard 3-lead ECG placement, with modified limb leads to measure the heart rate (HR)

#### 2. Multiple Fraction of Inspired Oxygen (FIO₂) Diffusing Capacity (DLCO) Method

1. Calculate the workloads corresponding to 30%, 50%, 70%, and 90% of the VO₂peak using the peak VO₂ obtained in the graded exercise test. At least 48 h after the graded exercise test, have the subject return to the laboratory for DLCO maneuvers.
2. Do not exceed 12 DLco tests per day, as carboxyhemoglobin (COHb) build-up can occur with repeated testing. Therefore, perform testing on multiple days based on the number of exercise workloads to be conducted and the quality of the DLCO data.
3. Prepare pre-breathing gases by attaching a tank of 100% O₂ gas and a tank of medical-grade air (21% O₂ and 79% N₂) to an air blender system. Fill two 60 L non-diffusing Douglas bags, one containing 40% O₂, and one containing 60% O₂, using the air blender system.
4. Set up two large-bore, three-way stopcock valves that will allow for the modulation of inhaled gas mixtures. These will be referred to as the "pre-breathing valves."
5. Connect the Douglas bags to the valve system using flexible, non-compressible tubing. Connect the valve system to a two-way, T-shaped non-re-breathing valve connected to the test gas intake assembly of the mass flow sensor of the metabolic measurement system.
6. For resting measurements, have the subject seated upright, with both feet flat on the floor. For exercise trials, ensure that the subject is in a steady state by monitoring HR using the ECG (HR ± 3 bpm for steady state).
7. Collect a single drop of capillary blood via a finger prick and analyze it for hemoglobin concentration. Then, adjust all subsequent DLCO for [Hb] using the following equation:

    \[
    DLCO_{adj} = DLCO \times \frac{10.22 + [Hb]}{1.7 \times [Hb]}
    \]
8. Select an FIO₂ (21%, 40%, or 60%) at random by switching the pre-breathe valves to the desired orientation. Choose the corresponding FIO₂ DLCO gas by turning the DLCO gas valve selector (see Figure 1C).
9. Instruct the subject to affix the nose clips and to breathe normally into the mouthpiece for five breaths from the Douglas bag corresponding to the respective FIO₂.
10. Instruct the subject to expire to residual volume. When the lung volume plateaus at residual volume, have the subject inhale the DLCO gas mixture to total lung capacity and hold their breath for 6 s before exhalation to residual volume.
11. Monitor the methane tracing during the exhalation to ensure that the slope is horizontal, as this indicates that the CO test gas is well equilibrated in the lung.
12. Ensure that the Vₐ for each DLco maneuver is within 5% of previous trials. Similarly, breath hold time should be 6.0 ± 0.3 s. If not, repeat the maneuver.
13. Wait 4 min to allow residual carbon monoxide to wash out, and then repeat steps 2.8 - 2.11 for each remaining FIO₂ at rest.
14. At least 48 h later, repeat steps 2.9 - 2.15 during steady state at each exercise intensity (30%, 50%, 70%, and 90% of the VO$_2$peak) for each F$_{O_2}$. Reduce the workload between the breath holds at 90% of the VO$_2$peak workload to recover the subject.

15. Wait 2 min between DLco tests during exercise to clear alveolar CO during exercise. Do not exceed 12 DLco tests per day to avoid carboxyhemoglobin (COHb) build-up$^5$.

3. Calculating Pulmonary Capillary Blood Volume and Membrane Diffusing Capacity

1. Calculate the alveolar partial pressure of O$_2$ (P$_A$O$_2$) using the following equation

$$P_aO_2 = F_{O_2}(P_{BAR} - P_{H2O}) - P_aCO_2 \times \frac{(1 - F_{O_2})}{RER}$$

NOTE: F$_{O_2}$ is the fraction of inspired O$_2$, P$_{BAR}$ is the atmospheric pressure, P$_{H2O}$ is the water vapor pressure, P$_aCO_2$ is the pressure of arterial CO$_2$, and RER is the respiratory exchange ratio.

2. Estimate the RER and P$_aCO_2$ using the measured 30-s average P$_{ET}CO_2$ and RER for the respective exercise intensity from the data obtained in the previous graded exercise test.

3. Calculate θ$_{CO}$ using the following equation$^7$,

$$\frac{1}{\theta_{CO}} = 0.0058 \times P_aO_2 + 0.73$$

4. Graph the relationship between 1/DLco$_{adj}$ and 1/θ$_{CO}$ for each F$_{O_2}$ and calculate the regression equation.

NOTE: The minimum acceptable r$^2$ value is 0.95, and DLco maneuvers should be repeated when r$^2$ values are outside of this range$^{21}$.

5. Calculate Vc by taking the inverse of the slope of the regression equation between 1/DLco$_{adj}$ and 1/θ$_{CO}$. Calculate Dm by taking the inverse of the y-intercept of the equation.

4. Intrapulmonary Arteriovenous Anastomosis Recruitment

1. On a separate day from the DLco data collection, insert a 20-gauge intravenous (IV) catheter into an antecubital vein and attach it to a three-way stopcock via a 6-in IV extension tube for the injection of agitated saline for contrast echocardiography$^{11,17}$. 
2. Connect two 10 mL syringes to the three-way stopcock. Combine 10 mL of 0.9% sterile saline with 0.5 mL of air, and forcefully agitate it through the three-way stopcock, back and forth between the two syringes, to form fine, suspended bubbles until the sonographer is ready for contrast.

3. Have an experienced sonographer or cardiologist obtain a standard apical four-chamber view of the heart. At rest, have the echocardiographer evaluate the intra-atrial septum and ventricular septum for an intra-cardiac shunt with standard echocardiographic and color Doppler imaging.
   
   1. If no intra-cardiac shunt is detected, instruct the subject to perform a Valsalva maneuver during the contrast injection to evaluate for a patent foramen ovale (PFO)\textsuperscript{11,17}. Repeat the measurement during non-Valsalva.

4. Inject the contrast while the sonographer maintains the four-chamber view. Record 15 cardiac cycles following the detection of contrast in the right ventricle.

5. Repeat the contrast-enhanced imaging during steady-state exercise at 30%, 50%, and 70% of the VO\textsubscript{2peak}. As steady state cannot be reached at 90% of the VO\textsubscript{2peak}, begin the imaging once the target HR, identified by the HR at 90% of the VO\textsubscript{2peak} during the graded exercise test, is reached.

   NOTE: The time between exercise intensities depends on the clearance of contrast from both ventricles, \( \geq 2 \) min.

6. Have an echocardiographer who is blinded to experimental conditions interpret the agitated saline contrast echocardiograms according to a previously-described scoring system\textsuperscript{17,27}.

   NOTE: Scoring is based on the maximum number of contrast bubbles visible within the left ventricle (LV) in a single echocardiographic frame, as follows: no contrast bubbles in the LV = 0, \( \leq 3 \) bubbles = 1, 4 - 12 bubbles = 2, > 12 bubbles = 3.

   NOTE: The appearance of contrast in the left ventricle after five cardiac cycles suggests an IPAVA. An intracardiac shunt is graded by the appearance of contrast in less than five cardiac cycles\textsuperscript{17}.

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**Representative Results**

The effect of increasing exercise intensity on oxygen consumption, diffusing capacity, pulmonary capillary blood volume, membrane diffusing capacity, and IPAVA score is shown in Table 1. VO\textsubscript{2}, DL\textsubscript{CO}, Vc, and Dm increase in response to increasing power output.

Figure 2 shows a representative calculation of Vc and Dm using the multiple F\textsubscript{I}O\textsubscript{2}-DL\textsubscript{CO} technique during exercise. DL\textsubscript{CO} decreases with increasing F\textsubscript{I}O\textsubscript{2}, and this relationship is exploited to partition Vc and Dm. Calculating the inverse of the slope of 1/DL\textsubscript{CO} versus 1/\( \theta \)\textsubscript{CO} results in the Vc, and the inverse of the y-intercept yields the value for the Dm. As expected, both the Vc and Dm increase during exercise compared to resting values.
The results show that these techniques can be used to assess the pulmonary vasculature response during exercise. The multiple-FI₂O₂ DL₉CO and agitated saline contrast echocardiography method provides investigators with more insight into the contributions of pulmonary capillary and membrane recruitment to the overall diffusion capacity and could supplement traditional pulmonary function testing in the clinical setting. Failure to increase Vc or Dm during exercise would lead to a diffusion limitation and hypoxemia. For example, a low DL₉CO secondary to a low Vc would indicate changes to the pulmonary capillaries; similarly, a decreased Dm would indicate changes to the pulmonary membrane.

Figure 4 shows representative tracings of four-chamber contrast echocardiographs. With increasing exercise intensity, the IPAVA score increases from 0 (i.e., no evidence of IPAVAs) at rest to 3 at the highest exercise intensity (Table 1). Previous work has shown that exercise increases the IPAVA score, but there is no consensus as to how these IPAVAs are recruited. There is evidence that IPAVAs can be recruited pharmacologically at rest with dopamine, as well as by increasing cardiac output with dobutamine and epinephrine. Inotropes such as dopamine and epinephrine are of particular interest, as they increase endogenously during exercise. Furthermore, there is some evidence that IPAVA recruitment may be important to exercise hemodynamics, in that the absence of IPAVAs appears to result in greater pulmonary artery pressure, decreased cardiac output, and decreased peak power output. Thus, this technique may be used in studies examining individuals with pulmonary artery hypertension.

**Discussion**

This method enables the evaluation of the pulmonary diffusing capacity and intrapulmonary arteriovenous anastomosis recruitment during exercise.
Critical steps within the protocol

Although the $DL_{CO}$ breath hold is relatively simple at rest, breath holding during exercise presents a unique challenge to the subject, as it is counter-intuitive, and subjects have a high drive to breathe during exercise. Thus, a good-quality determination of $Vc$ and $Dm$ relies on the rapport and clear communication between the tester and the subject. The tester’s technical ability can be quantified with the variability of the alveolar volume (± 5% of previous trials) and a breath-hold time (BHT) of 6.0 ± 0.3 s.

Modifications and troubleshooting

At the conclusion of a $Vc/Dm$ measurement, the tester should quickly graph the three $DL_{CO}$ maneuvers to determine the best-fit line of the data points; the $DL_{CO}$ measured with 21% $F_O2$ should always be greater than that with 40%, which should be greater than that with 60%. If not, it is recommended to check if the valve switch corresponds to the correct testing gas. Similarly, check that the pre-breathing bags are filled with the correct $F_O2$ gas corresponding to the testing gas (Figure 1B-1D). Caution should be taken when testing a participant who is a smoker, as elevated COHb levels may underestimate $DLco$.

For the IPAVA recruitment assessment, the position of the subject is critical to ensure high-quality image acquisition. It is possible to replace the upright cycle ergometer with a recumbent cycle ergometer to minimize the movement of the subject. However, recumbent cycle exercise will elicit a different metabolic response for a given work rate, and thus the graded exercise test should be repeated on the recumbent cycle ergometer. Scanning of the upper chest may be uncomfortable to some women; in this case, a female sonographer is recommended. Finally, the recommended exercise protocol is designed for a young, healthy individual; accordingly, the exercise protocol can be modified for a different target population.

Limitations of the technique

The principal limitations of the multiple $F_O2$ $DL_{CO}$ technique are the skill of the tester and the ability of the subject to follow commands and to remain calm during the breath hold, as Valsalva or Müllerian maneuvers will affect the measurements. Secondly, the number of breath holds in one session should be limited to 12, due to an increase in CO backpressure, which may affect the $Vc$ and $Dm$ measurement and pose a health risk to the subject. Depending on the research design, it may be necessary to complete the testing across multiple sessions to allow for the clearance of CO and to limit participant fatigue. With good participant coaching and good technical ability, we have determined a satisfactory coefficient of variation between trials for $DLco$, $Vc$, and $Dm$ to be 7%, 8%, and 15%, respectively.

The multiple $F_O2$ $DL_{CO}$ technique assumes that the alveolar $O_2$ is the same as the capillary $O_2$, and thus, caution should be exercised when interpreting the data in individuals with known gas exchange impairment.

Agitated saline contrast echocardiographic imaging is limited by the technical ability of the sonographer and the ability of the subject to minimize thoracic movement while exercising. It is also critical that the interpreter of the images be familiar with the scale for scoring IPAVA recruitment according to established procedures (Figure 4). The significance of a positive saline contrast echocardiography during exercise remains a topic of debate, and there is some discussion that a positive agitated saline contrast in the left ventricle may be secondary to capillary distention, and not IPAVA recruitment. Ongoing work is attempting to resolve this issue.

Significance of the technique with respect to existing/alternative methods

By utilizing these physiological techniques, it is possible to assess the pulmonary vasculature during exercise in a variety of conditions, including in health, in disease, and in drug interventions. Although the quality relies with the ability of the tester, these skills are easily and quickly acquired with proper mentorship and training. The multiple $F_O2$ $DL_{CO}$ method is considered the "gold standard" in the measurement of $Dm$ and $Vc$. While these measures are not calculated clinically, the values could be used to determine the mechanisms for hypoxemia and exercise intolerance, to predict patient outcomes, and to further characterize diagnosis. Likewise, the agitated saline echocardiography technique is the most widely-used method in determining the recruitment of IPAVAs.

Future applications or directions after mastering this technique

These techniques are applicable for use in a range of experimental conditions and interventions. We demonstrate these techniques during exercise, but they can easily be modified to measure pulmonary vascular responses during a drug infusion, such as dobutamine or dopamine, inotropes known to increase cardiac output. Furthermore, it is possible to use these techniques in clinical populations, such as those with heart failure or chronic obstructive pulmonary disease (COPD), in which the $DL_{CO}$ is lower compared to age-matched control subjects.

Disclosures

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